#### Review Article

# MicroRNAs in clear cell renal cell carcinoma: biological functions and applications

# Gianluca Aguiari

Department of Biomedical and Surgical Specialty Sciences, Section of Biochemistry, Molecular Biology and Medical Genetics, University of Ferrara, Italy.

### **Abstract**

MicroRNAs (miRs) are small noncoding RNAs that govern many biological processes. They frequently acquire a gain or a loss of function in cancer and hence play a causative role in the development and progression of neoplasms. They could be used as biomarkers to improve our knowledge on diagnosis, prognosis and drug resistance, and to attempt therapeutic approaches in several types of cancer including clear cell renal cell carcinoma (ccRCC). ccRCC is the most predominant subtype of RCC that accounts for about 90% of all renal cancers. Since ccRCC is generally asymptomatic until very late, it is difficult to diagnose early. Moreover, in the absence of preventive treatments for metastatic ccRCC after surgical resection of the primary cancer, predictive prognostic biomarkers are needed in order to achieve appropriate therapies. Herein the role of miRs in the biology of ccRCC and the potential applications of these molecules are discussed. Moreover, future applications in the diagnostic and prognostic field, as well as their impact on drug response and therapeutic targets are also explored. Their use in clinical practice as molecular biomarkers alone, or in combination with other biological markers could accelerate progress, help design personalized therapies, limit side effects, and improve quality of life of ccRCC patients.

Received: 16 July 2015; Accepted after revision: 18 August 2015; Published: 23 August 2015.

**Author for correspondence:** Gianluca Aguiari, Department of Biomedical and Surgical Specialty Sciences, Section of Biochemistry, Molecular Biology and Medical Genetics, University of Ferrara, Italy. E-mail: dsn@unife.it

**How to cite:** Aguiari G. MicroRNAs in clear cell renal cell carcinoma: biological functions and applications. Journal of Kidney Cancer and VHL 2015;2(4):140-152. Doi: http://dx.doi.org/10.15586/jkcvhl.2015.40

### Introduction

Renal cell carcinoma (RCC) accounts for 2-3% of all cancers and is the most common kidney malignancy with the highest mortality rate of urinary cancers. Lifestyle factors including smoking, obesity and hypertension are considered etiological risk factors for this tumor (1-3). RCC is mainly classified into three major subtypes: clear cell (ccRCC), papillary (pRCC) and chromophobe (chRCC). The ccRCC subtype is the most common accounting for 80-90% of RCC cases (3). Generally, ccRCC arises

from the epithelial cells of the proximal convoluted renal tubule and is histologically characterized by clear cells separated by hypervascular thin fibrous septae (4-5). This subtype of renal cancer commonly metastasizes to the bones, lungs, brain and liver through the invasion of the vena cava Currently, partial or radical nephrectomy is the only curative treatment for localized tumors with high quality of life outcomes (3). In comparison, for patients with metastatic ccRCC, nephrectomy is mostly palliative and systemic treatment with pharmacological therapy is necessary.

Unfortunately, ccRCC is unresponsive to traditional chemotherapies, highly resistant to radiation, and lacks the hallmark genetic features of solid tumors, such as mutations in KRAS and TP53 genes (7).

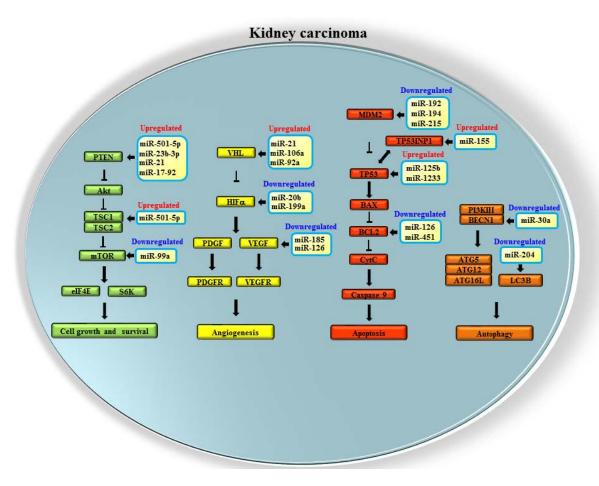
Recent advances in molecular biology have led to the identification of a plethora of gene mutations associated with the development and progression of ccRCC. It is well known that the von Hippel-Lindau (VHL) gene, localized on chromosome 3p, is frequently either inactivated by mutation methylation in over 80% of ccRCC patients. The VHL protein complexes with other proteins, and functions as an E3 ubiquitin ligase. This leads to degradation of the a subunit of hypoxia-inducible transcription factors (HIF1 and HIF2) via proteasome activation (8-9). The loss of function of VHL prevents the degradation of HIFa proteins resulting in the increased expression of angiogenic factors including vascular endothelial growth factor (VEGF) and platelet-derived growth factor B chain (PDGF-B) that contribute to growth and expansion of tumor (10). Mutations in mammalian target of rapamycin (mTOR), TSC1, PIK3CA, and PTEN genes in approximately 20% of ccRCC have also been observed. These molecular lesions cause an aberrant mTORC1 pathway activation that could play a relevant role in the neoplastic transformation of kidney cells (8). The discovery of several altered signaling pathways associated with ccRCC has opened new opportunities and strategies in the treatment of ccRCC, especially the targeted inhibition of molecules involved in VEGF and mTOR pathways (11-12). However, complete remission or long-term beneficial responses with tyrosine kinase or mTOR inhibitors are rare (12). Moreover, these pharmacological approaches are expensive and show significant adverse effects that worsen the quality of life of ccRCC patients. Thus, cost-effective predictive biomarkers that are able to improve clinical management of ccRCC patients are needed. In this regard, an attractive solution could be offered by microRNAs (miRs) that are involved in the development of many cancers. Herein we discuss the role of miRs in RCC biology and their possible applications as biomarkers for ccRCC.

### Biogenesis and function of microRNAs

MiRs are short non-coding single stranded RNAs of 20-22 nucleotides in length that regulate gene expression at the posttranscriptional level. They act by inhibiting the translation of mRNA primarily by pairing with the 3'-untranslated region (UTR) of their complementary mRNAs (13). MiRs are transcribed by RNA polymerase II or RNA polymerase III into a long primary nuclear miR (primiRNA) that is cleaved to 70-80 bp long pre-miRNA by the Drosha/DGCR8 complex and exported into the cytoplasm via exportin-5/RanGTP (14-15). Pre-miRNAs are then digested to double-stranded mature miRs by the RNAse III Dicer, and separated in two strands: the miRNA-guide strand and the miRNA-passenger strand. The miRNA-guide strand is loaded into the RNA-induced silencing complex (RISC) and driven to the target mRNA, while the miRNApassenger strand is generally degraded. MiRNA gene silencing may be performed through degradation via complete pairing, where the mRNA target is degraded; or incomplete pairing, with the inhibition of translation or the decay of mRNA into cytoplasmic foci termed P bodies (14). MiRs regulate several cellular processes such as apoptosis, proliferation, hematopoiesis and angiogenesis (13). Pertinent to tumors, they can act either as pro- or anti-oncogenic molecules, thereby having a dual role in oncogenesis. They can act either as tumor suppressors by targeting oncoproteins or as tumor promoters by downregulating the expression of tumor suppressor proteins (16-17). Moreover, a large number of miRs have been identified to regulate cellular metabolism that is often altered in tumor cells contributing to development and progression of cancer (18).

# MicroRNAs affect biological processes in ccRCC

It is well known that miRs are involved in the pathophysiology of a variety of kidney diseases including diabetic nephropathy, polycystic kidney disease, Wilms tumor and kidney cancer (13). This section focuses on the role of miRs in kidney cancer with emphasis on cell growth, angiogenesis, apoptosis and autophagy (**Figure 1**; **Table 1**).



**Figure 1:** Dysregulated microRNAs affecting cell growth, angiogenesis, apoptosis and autophagy in kidney cancer. In the sketch, microRNAs targeting genes involved in mTOR (green), VHL/HIF-a (yellow) and p53 (red) pathways as well as in autophagic process (orange) are shown.

### Cell growth and migration

As observed in other tumors, miRs may contribute to development of RCC by modulating the expression of genes involved in this cancer. For example, VHL is targeted by miR-106a, miR-92a and miR-21, which have been found upregulated in renal carcinoma tissues (19-20). Moreover, VHL loss of function can activate the expression of different miRs including the miR-17-92 cluster, also known as oncomiR-1. These miRs act as oncogenic agents targeting PTEN and E2Fs tumor suppressor genes Furthermore, **VHL** (21-22).deletion/mutation also causes the enhancing of miR-28-5p expression that in turn induces the reduction of Mad2 protein levels, thereby promoting checkpoint weakness and chromosomal instability in

ccRCC cells and tissues (23). In addition, the impaired VHL signaling causes the activation of HIF pathway, which leads to the upregulation of miR-210 and miR-155 which have been identified as having oncogenic properties in ccRCC (19-20, 24). Conversely, hypoxia can lead to the downregulation of miRs that target the HIF1a transcripts. MiR-20b and miR-199a are repressed under conditions of hypoxia (25). Furthermore, hypoxia may reduce miRs that target VEGF and promote angiogenesis (25). Besides VHL gene, miR-21 may degrade PTEN mRNA leading to the activation of Akt and mTOR kinases which contribute to growth and migration of kidney cancer cells (26). In ACHN and 786-0 cells, miR-21 induced migration and invasion through the modulation of the proapoptotic protein PDCD4 (19, 27).

Table 1: MicroRNAs dysregulated in renal cancer

MicroRNAs	Target genes	Expression	Role	Applications	Ref.
miR-501-5p, miR- 23b-3p	TSC1, PTEN, MCU	Up	Cell growth, metastasis	Prognosis	2, 28
miR-21, 106a, 92a	VHL, PTEN, PDC4	Up	Cell growth, migration	Prognosis	19-20, 26- 27, 52
miR-210, 155	ISCU1/2, TP53INP1	Up	Metastasis, cell growth	Diagnosis/prognosis	20, 24, 44- 45, 49-50
miR-20b, 199a	HIF-a	Down	Anti-angiogenesis	-	25
17-92 cluster	PTEN, E2Fs	Up	Cell growth, invasion	Prognosis	20-22, 24
miR-28-5p	Mad2	Up	Chromosomal instability	-	23
miR-192, 194, 215	ZEB2, MDM2, TYMS	Down	Cell differentiation	Prognosis	29
miR-584	ROCK1	Down	Cell cycle arrest	-	30
miR-135a	cMYC	Down	Cell cycle arrest	-	31
miR-205	SFKs	Down	Cell cycle arrest, apoptosis	-	32
miR-708	Survivin, ZEB2, BMI1	Down	Apoptosis	-	33
miR-1826	CTNNB1, MEK1	Down	Cell cycle arrest, apoptosis	-	34
miR-30d, 185	MTDH, VEGFA	Down	Apoptosis	-	35-36
miR-204	LC3B	Down	Autophagy	-	37-38
miR-30a	BECN1	Down	Inhibition of autophagy	-	39
miR-203, 424	FGF2, PDCD4	Up	Cell proliferation	Diagnosis	41
miR-195, 15b	Raf-1, CCND1	Up	Cell cycle arrest	Diagnosis	42
miR-122-5p	HIF1- a vimentin	Up	Inhibition of cell invasion	Diagnosis	43
miR-378, 451	TOB2, BCL2	Up/down	Apoptosis, cell cycle	Early diagnosis	46
miR-1233	BLCAP, TP53	Up	Inhibition of apoptosis	Early diagnosis	47
miR-193a, 362, 572, 28, 378	MAPK, NFkB, Mad2	Up/down	Cell growth and invasion	Early diagnosis	48
miR-126	VEGFA, BCL2	Down	Cell migration, apoptosis	Prognosis	51-52
miR-125b	E2F3, TP53	Up	Cell growth and migration	Prognosis	53
miR-30c	Snail1/slug, PTP4A1	Down	Inhibition of cell migration	Prognosis	55
miR-200c	CYP1B1, HO-1	Down	Inhibition of cell invasion	Drug response	58-59
miR-942	BRCA pathway	Up	Angiogenesis, metastasis	Drug response	60
miR-141	ZEB2	Down	Cell differentiation	Drug response	61
miR-200	IL-8, CXCL1	Down	Anti-angiogenesis	Therapy	63
miR-99a	mTOR	Down	Cell cycle arrest	Therapy	64

In addition, it also promoted tumor metastasis by negatively regulating the

tumor suppressor gene tropomyosin-1 (19). Other miRs targeting PTEN and mTOR

mRNAs such as miR-501-5p and miR-23b-3p show oncogenic features in different ccRCC cell lines (2, 28). In ccRCC tissue of patients that developed distant metastases, a significant increase of miR-501-5p expression was observed (2). In KJ29 and Caki-2 ccRCC cells, miR-501-5p stimulated cell proliferation via mTOR-mediated MDM2 upregulation. MiR-23b-3p was upregulated in A-498 and Caki-2 ccRCC cell lines and its downregulation induced apoptosis and reduced invasion (28). In patients, an increased expression correlated with lower survival rate (28).

# Cell differentiation and apoptosis

The downregulation of miRs targeting specific oncogenes may contribute to the development and progression of RCC. In this regard, miR-192, miR-194 and miR-215 are highly expressed in normal kidney, but they are significantly downregulated in both primary and metastatic ccRCC. These miRs target ZEB2, MDM2 and TYMS mRNAs that are associated with cancer development and metastasis. The restoration of these miRs causes a reduction of cell migration and invasion in RCC cell lines (29). Moreover, miR-584, which is downregulated in ccRCC tissues, functions as a tumor suppressor miR that inhibits cell motility by the negative regulation of ROCK-1 oncogene expression (30). Similarly, miR-135a was found significantly reduced in renal cancer tissues and its restoration in both Caki-2 and A498 ccRCC cell lines caused cell cycle arrest in G0/G1 phase. It also targeted the c-MYC oncogene which was upregulated in ccRCC tissues (31). Cell cycle arrest may be also induced by miR-205 restoration that leads to inhibition of Src family kinases (SFKs) expression, stimulating apoptosis and inhibiting cell proliferation, migration, and colony formation in renal cancer cells. This miR is significantly suppressed in renal cancer cell lines and tumor tissues (32).

Pro-apoptotic miRs are often downregulated in ccRCC. For instance, miR-708, which exerts pro-apoptotic effects in ccRCC cells and tissues by targeting survivin, ZEB2 and BMI1 genes that are involved in cell survival, adhesion, invasion, and metastasis, has been found widely reduced in human RCC tissues. The restoration of miR-708 dramatically increases apoptosis in ccRCC

cell lines. Moreover, the intra-tumoral of this miR stimulates delivery regression of tumors in murine xenograft models of human RCC (33). Similarly, miR-1826 that is significantly downregulated in renal cancer tissues acts as tumor suppressor agent promoting apoptosis and cell cycle arrest by knockdown of CTNNB1 (beta-catenin) and MEK1 genes in VHLinactivated ccRCC cells (34). The miRs miR-30d and miR-185 also may function as onco-suppressors in ccRCC cells inducing apoptosis through the inhibition of the oncoprotein metadherin (MTDH) and VEGF A (VEGFA) expression (35-36).

## Autophagy

Autophagy is an important mechanism that plays both pro- and anti-oncogenic roles. In kidney cancer, the pro- or anti-oncogenic role of autophagy is mediated by two different forms of autophagic protein LC3, LC3B and C, which are regulated by the VHL protein. The expression of LC3B supports tumor formation and progression in renal cancer cells with VHL loss of function, whilst the expression of LC3C shows tumor suppressor activity (37). A functional VHL protein inhibits LC3B-mediated autophagy through the stimulation of miR-204, which directly targets LC3B mRNA reducing its translation. In ccRCC cells and metastatic tissues, most of which are characterized by early inactivation of the VHL gene, a reduction of miR-204 expression as well as an increase of LC3B protein levels have been reported. Thus, in kidney cells, miR-204, in a network involving VHL, functions as a tumor suppressor miR that inhibits prooncogenic autophagy by targeting LC3B mRNA (37-38). The expression of miR-30a may also affect autophagy in kidney cancer. It acts as an onco-suppressing agent and inhibits autophagy by targeting the BECN1 gene that codes for beclin-1, a protein formation crucial for the autophagosome. This miR is significantly reduced in both human RCC tissues and cell lines, and its restoration not only causes the inhibition of autophagy by the reduction of beclin-1 protein but also enhances sorafenib-induced cytotoxicity in different ccRCC cell lines (39). Autophagy may represent an attractive target for the design of new therapeutic approaches in chemoresistant metastatic renal cancer by using specific drugs for the inhibition of prooncogenic autophagy. However, further studies are needed to evaluate the pro- or anti-oncogenic role of autophagy in renal cancer.

# MicroRNAs and their applications in ccRCC

In RCC patients after surgical resection no effective preventive adjuvant therapy is available. The likelihood of relapse or metastases in high grade ccRCC is frequent Moreover, the lack of specific prevents prognostic biomarkers development of specific therapy. Thus, the discovery of new biomarkers for prognosis of ccRCC could be useful for the clinical management of these cancers (2). In this regard, miRs that are involved carcinogenesis could be considered as predictive biomarkers for kidney cancer. The stable nature of these molecules and their relative ease of detection and recovery in fluids biological make them optimal candidates as molecular biomarkers. In particular, they could be used diagnostic/prognostic tools and for the design of new therapeutic strategies (Figure

# MicroRNAs for the identification of kidney cancer subtypes

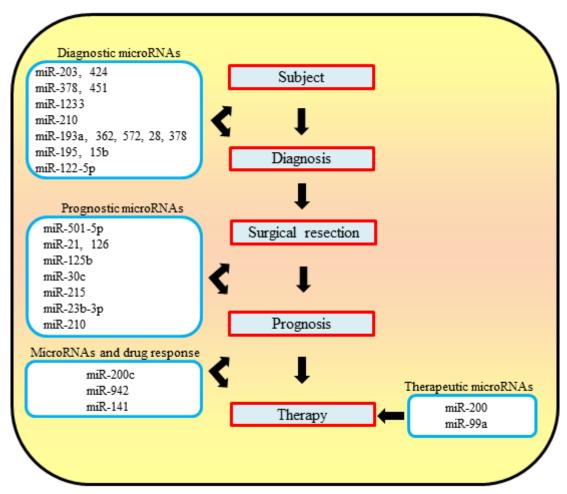
Immunohistochemical markers do not allow accurate identification of subtypes, especially in the setting of uncommon morphology and small biopsies. However, miRs that are well established as regulators of tumorigenesis could be utilized as potential biomarkers alone or in combination with current markers in order to improve the classification of kidney cancer. In this regard, miR-424 and miR-203 that are overexpressed in ccRCC but not in pRCC could be used to distinguish ccRCC from pRCC (41). Also miR-195 and miR15b are upregulated ccRCC in chRCC compared with pRCC, oncocytoma (42). The expression of miR-126-3p is higher in ccRCC than in pRCC tissues, while miR-122-5p, which is highly expressed in liver, was specifically expressed in ccRCC but not in the other subtypes of RCC (43). The differential expression of 24 miRs in different kidney cancer subtypes was reported to distinguish all 4 common subtypes of kidney tumors with an overall accuracy of 95% (43). Therefore, these miRs could be considered as diagnostic biomarkers in order to improve the classification of renal cell carcinoma.

# Circulating microRNAs for early diagnosis of ccRCC

Circulating miRs could serve as noninvasive biomarkers for the early diagnosis of kidney carcinoma. In this regard, miR-210 that is strongly upregulated in ccRCC tissue, is also abundantly found in the serum of ccRCC patients compared with healthy controls. These findings suggest that miR-210 could be used as biomarker for early diagnosis of ccRCC (19,44-45). Other studies report that levels of miR-378 are increased, while those of miR-451 are decreased in serum of RCC patients healthy controls. compared with detection of miR-378 and miR-451 in serum allows the identification of patients with RCC with a sensitivity of 81% and a specificity of 83%, making the combination of these miRs a possible early diagnostic biomarker for RCC (46). Also the circulating miR-1233 could also serve as a potential diagnostic biomarker for this cancer (47). Recently, it has been reported that serum levels of miR-193a-3p, miR-362 and miR-572 are significantly increased whereas those of miR-28-5p and miR-378 are decreased in patients with RCC compared with the healthy controls. The combined use of these 5 miRs could serve as a biomarker panel for the identification of early stage renal cancer with a sensitivity of 80% and a specificity of 71% (48). However, further studies are needed to validate candidate circulating miRs as biomarkers for early diagnosis of renal cancer by using large multicentre-cohorts of patients.

## *MicroRNAs for ccRCC prognosis*

Apart from early diagnosis, miRs could also serve as prognostic markers. In this regard, a high level of miR-210 was associated with a good clinico-pathological outcome in



**Figure 2**: Flow chart of possible future management of RCC patients by using microRNAs. MicroRNAs with diagnostic, prognostic and therapeutic properties along with drug response are indicated. Prognostic miRs should allow the identification of ccRCC patients that need specific therapeutic treatments from those which do not need pharmacological therapy.

ccRCC patients (49), however, other studies have associated a high miR-210 with a higher chance of disease recurrence and shorter overall survival (24, 50). This discrepancy suggests that further studies to define the role of miR-210 in ccRCC are needed. Lower levels of miR-215 (29) and miR-126 (51) are associated with a worse outcome in RCC patient and a higher expression of miR-126 is associated with significantly prolonged disease-free survival and overall survival (51). Furthermore, an increased miR-501-5p, miR-23b-3p (2, 28), miR-21 (52), miR-125b (53) and a decreased miR-126 (52) are indicators of worse outcome in metastatic patients. Other studies report that the high miR-21/10b ratio is associated with a poor prognosis in ccRCC non-metastatic patients Recently, it has been published that the differential expression of miR-30c, miR-451 and miR-126 are associated with overall survival in ccRCC patients (55). A miR signature that includes the upregulation of miR-21-5p, 142-3p, let-7g-5p, let-7i-5p and 424-5p, as well as the downregulation of miR-204-5p is associated with high stage, grade and progression of ccRCC (56). Another miRNA signature consisting of 22 miRNAs that are significantly linked to patient survival in ccRCC was recently identified (57). Finally, it has also been reported that the expression some miRs may predict site of metastasis. For example, the decreased expression of miR-10b associated with brain metastasis, while the upregulation of miR-199b and downregulation of miR-615 are linked to lung metastasis in patients with advanced ccRCC (55).

### *MicroRNAs and drug resistance*

In addition to their role in tumor progression, miRs could also affect response to therapy. The downregulation of miR-200c in RCC causes increased expression of the CYP1B1 gene resulting in pharmacological resistance to docetaxel treatment (58). Moreover, the upregulation of miR-200c enhanced the sensitivity of ccRCC cells to sorafenib or imatinib, inhibiting proliferation by targeting heme oxygenase-1 (HO-1) mRNA (59). The upregulation of miR-942, miR-133a, miR-628-5p, and miR-484 was associated with sunitinib resistance in metastatic RCC patients. In particular, the increased expression of miR-942 alone could accurately predict sunitinib MMP-9 resistance that occurs by upregulation and VEGF secretion (60). studies show that miR-141 epithelial-todownregulation drives mesenchymal transition (EMT) in ccRCC and is associated with an unfavorable response to sunitinib treatment. In vitro reintroduction of miR-141 reversed EMT and decreased cell viability in hypoxic conditions (61). To date, about 28 miRNAs related to poor response and 23 associated with prolonged response to sunitinib treatment have been identified (62).

### *MicroRNAs for gene therapy*

As previously discussed, miRs may act as onco-suppressors, or contributing to development, growth and metastasis of RCC. Therefore, the restoring of normal levels of onco-suppressor miRs or the degradation of onco-miRs by using specific anti-miR sequences could be an attractive therapeutic strategy for the treatment of metastatic ccRCC. In this regard, the downregulation of the miR-200 family in different tumors is associated with cell migration, invasion, and metastasis by the increased expression of IL-8 and CXCL1 genes that promote angiogenesis. The delivery of miR-200 members into the tumor endothelium of orthotopic cancer models by using specific nanoparticle resulted in marked reduction of metastasis and angiogenesis (63). The downregulation of miR-99a in RCC tissues was associated with distant metastases and poor prognosis in RCC patients through the activation of the mTOR pathway. The restoration of miR-99a levels in RCC cells inhibited cell growth, migration and invasion. Moreover, intratumoral delivery of miR-99a reduced tumor growth in murine xenograft models of human RCC (64). The development of specific miR-targeted therapies, such as the reintroduction of onco-suppressor miRNAs or the inhibition of onco-miRs in RCC cells, could have significant translational implications such as improvement of the clinical picture, management and quality of life of RCC patients. However, there are several obstacles to deliver miRs/anti-miRs into the tumor cell including low cellular uptake, endosomal escape, immunogenicity, degradation in the bloodstream, and rapid renal clearance. Moreover, for a successful miR-based therapy, specific delivery of miR molecules to target the organ is important. There is an need for developing efficient transfection methods as well as targetspecific delivery systems to realize the full therapeutic potential of miRs (65).

#### **Conclusions**

This review gives an overview of the impact of miRs in renal tumorigenesis, their potential role as biomarkers and therapeutic targets. MiRs show a relative versatility of use, however they may also show some practical disadvantages largely because of with associated problems accurate quantification in biological fluids. In fact, miRNA analysis in paraffin embedded tissues as well as in serum or in other biological fluids may be complicated by the specific methods absence of standardization and suitable cut-off values. Furthermore, miR expression could be affected by genetic factors, environment, lifestyle and by sample collection methods. Despite these potential limitations, a large number of studies showing miRs as predictive biomarkers in different cancer types are emerging. Further studies using large independent cohorts of ccRCC patients are needed to validate miRs as biomarkers that are useful in clinical practice. The combination of miRs and other renal cancer biomarkers such as VEGF and VEGFrelated proteins, cytokines and lactate dehydrogenase (66) could improve the reliability of these putative biomarkers. Therefore, miRs could become a powerful tool for clinical management of high grade

and metastatic ccRCC patients, opening new horizons to achieve personalized medicine.

### **Acknowledgements**

We kindly thank Dr. Lucio Dell'atti of Urology Unit (Sant'Anna Hospital of Ferrara, Italy) for support on article drafting. This work is supported by Italian Ministry of Health (Ricerca Finalizzata 2011-2012) grant no. GR-2011-02346964, University of Ferrara local funds: FAR 2012 and 2013.

#### Conflict of interest

None

#### References

- 1. Cairns P. Renal cell carcinoma. Cancer Biomark. 2010:9:461-473. PMid:22112490.
- 2. Mangolini A, Bonon A, Volinia S, Lanza G, Gambari R, Pinton P, Russo GR, Del Senno L, Dell'Atti L, Aguiari G. Differential expression of microRNA501-5p affects the aggressiveness of clear cell renal carcinoma. FEBS Open Bio. 2014;4:952-65.

http://dx.doi.org/10.1016/j.fob.2014.10.0 16

PMid:25426415 PMCid:PMC4241533.

3. Ljungberg B et al. EAU Guidelines on Renal Cell Carcinoma: 2014 Update. Eur Urol. 2015;67(5):913-24.

http://dx.doi.org/10.1016/j.eururo.2015.0 1.005

PMid:25616710.

4. Baldewijns MM, van Vlodrop IJ, Schouten LJ, Soetekouw PM, de Bruïne AP, van Engeland M. Genetics and epigenetics of renal cell cancer. Biochim Biophys Acta. 2008;1785(2):133-55.

http://dx.doi.org/10.1016/j.bbcan.2007.1 2.002

5. Montironi R, Mazzucchelli R, Scarpelli M, Lopez-Beltran A, Cheng L. Update on selected renal cell tumors with clear cell features. With emphasis on multilocular cystic clear cell renal cell carcinoma. Histol Histopathol. 2013;28(12):1555-66. PMid:23893851.

6. Milowsky MI, Nanus DM. Chemotherapeutic strategies for renal cell carcinoma. Urol Clin North Am 2003;30:601-609.

http://dx.doi.org/10.1016/S0094-0143(03)00031-4

7. Jonasch E et al. State of the science: an update on renal cell carcinoma. Mol Cancer Res. 2012;10(7):859-80.

http://dx.doi.org/10.1158/1541-7786.MCR-12-0117

PMid:22638109 PMCid:PMC3399969.

8. Brugarolas J. Molecular genetics of clearcell renal cell carcinoma. J Clin Oncol. 2014;32(18):1968-76.

http://dx.doi.org/10.1200/JCO.2012.45.2 003

PMid:24821879 PMCid:PMC4050206.

9. Gossage L, Eisen T, Maher ER. VHL, the story of a tumour suppressor gene. Nat Rev Cancer. 2015;15(1):55-64.

http://dx.doi.org/10.1038/nrc3844 PMid:25533676.

10. Gudas LJ, Fu L, Minton DR, Mongan NP, Nanus DM. The role of HIF1a in renal cell carcinoma tumorigenesis. J Mol Med (Berl). 2014;92(8):825-36.

http://dx.doi.org/10.1007/s00109-014-1180-z

PMid:24916472 PMCid:PMC4119538.

11. Audenet F, Yates DR, Cancel-Tassin G, Cussenot O, Rouprêt M. Genetic pathways involved in carcinogenesis of clear cell renal genomics towards cell carcinoma: personalized medicine. BJU Int. 2012;109(12):1864-70.

http://dx.doi.org/10.1111/j.1464-410X.2011.10661.x PMid:22035299.

12. Srinivasan R, Ricketts CJ, Sourbier C, Linehan WM. New strategies in renal cell carcinoma: targeting the genetic and metabolic basis of disease. Clin Cancer Res. 2015;21(1):10-7.

http://dx.doi.org/10.1158/1078-0432.CCR-13-2993

PMid:25564569.

13. Badal SS, Danesh FR. MicroRNAs and their applications in kidney diseases. Pediatr Nephrol. 2015;30(5):727-40.

http://dx.doi.org/10.1007/s00467-014-2867-7

PMid:24928414.

14. Romero-Cordoba SL, Salido-Guadarrama I, Rodriguez-Dorantes M, Hidalgo-Miranda A. miRNA biogenesis: biological impact in the development of cancer. Cancer Biol Ther. 2014;15(11):1444-55.

http://dx.doi.org/10.4161/15384047.2014 .955442

PMid:25482951.

15. Yi R, Qin Y, Macara IG, Cullen BR. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. Genes Dev. 2003;17:3011-6.

http://dx.doi.org/10.1101/gad.1158803 PMid:14681208 PMCid:PMC305252.

16. Reddy KB. MicroRNA (miRNA) in cancer. Cancer Cell Int. 2015;15:38.

http://dx.doi.org/10.1186/s12935-015-0185-1

PMid:25960691 PMCid:PMC4424445.

17. Iorio MV, Croce CM. Causes and consequences of microRNA dysregulation. Cancer J. 2012; 18(3):215-22.

http://dx.doi.org/10.1097/PPO.0b013e31 8250c001

PMid:22647357 PMCid:PMC3528102.

18. Chan B, Manley J, Lee J, Singh SR. The emerging roles of microRNAs in cancer metabolism. Cancer Lett. 2015;356(2 Pt A):301-8.

http://dx.doi.org/10.1016/j.canlet.2014.1 0.011

PMid:25451319.

19. Chow TF, Youssef YM, Lianidou E, Romaschin AD, Honey RJ, Stewart R, Pace KT, Yousef GM. Differential expression profiling of microRNAs and their potential involvement in renal cell carcinoma pathogenesis. Clin Biochem. 2010;43(1-2):150-8.

http://dx.doi.org/10.1016/j.clinbiochem.2 009.07.020

PMid:19646430.

20. Valera VA, Walter BA, Linehan WM, Merino MJ. Regulatory Effects of microRNA-92 (miR-92) on VHL Gene Expression and the Hypoxic Activation of miR-210 in Clear

Cell Renal Cell Carcinoma. J Cancer. 2011;2:515-26.

http://dx.doi.org/10.7150/jca.2.515 PMid:22043236 PMCid:PMC3204400.

21. Chow TF et al. The miR-17-92 cluster is over expressed in and has an oncogenic effect on renal cell carcinoma. J Urol. 2010;183(2):743-51.

http://dx.doi.org/10.1016/j.juro.2009.09. 086

PMid:20022054.

22. Mogilyansky E, Rigoutsos I. The miR-17/92 cluster: a comprehensive update on its genomics, genetics, functions and increasingly important and numerous roles in health and disease. Cell Death Differ. 2013;20(12):1603-14.

http://dx.doi.org/10.1038/cdd.2013.125 PMid:24212931 PMCid:PMC3824591.

23. Hell MP, Thoma CR, Fankhauser N, Christinat Y, Weber TC, Krek W. miR-28-5p promotes chromosomal instability in VHL-associated cancers by inhibiting Mad2 translation. Cancer Res. 2014;74(9):2432-43

http://dx.doi.org/10.1158/0008-5472.CAN-13-2041 PMid:24491803.

24. Neal CS, Michael MZ, Rawlings LH, Van der Hoek MB, Gleadle JM. The VHL-dependent regulation of microRNAs in renal cancer. BMC Med. 2010;8:64.

http://dx.doi.org/10.1186/1741-7015-8-64

PMid:20964835 PMCid:PMC2978113.

25. Nallamshetty S, Chan SY, Loscalzo J. Hypoxia: a master regulator of microRNA biogenesis and activity. Free Radic Biol Med. 2013;64:20-30.

http://dx.doi.org/10.1016/j.freeradbiomed .2013.05.022

PMid:23712003 PMCid:PMC3762925.

26. Dey N, Das F, Ghosh-Choudhury N, Mandal CC, Parekh DJ, Block K, Kasinath BS, Abboud HE, Choudhury GG. microRNA-21 governs TORC1 activation in renal cancer cell proliferation and invasion. PLoS One. 2012;7(6):e37366.

http://dx.doi.org/10.1371/journal.pone.00 37366

PMid:22685542 PMCid:PMC3368259.

27. Bera A, Das F, Ghosh-Choudhury N, Kasinath BS, Abboud HE, Choudhury GG. microRNA-21-induced dissociation of PDCD4 from rictor contributes to Akt-IKKβ-mTORC1 axis to regulate renal cancer cell invasion. Exp Cell Res. 2014;328(1):99-117. http://dx.doi.org/10.1016/j.yexcr.2014.06.022

PMid:25016284.

28. Zaman MS et al. Inhibition of PTEN gene expression by oncogenic miR-23b-3p in renal cancer. PLoS One. 2012;7(11):e50203. http://dx.doi.org/10.1371/journal.pone.00 50203

PMid:23189187 PMCid:PMC3506541.

29. Khella HW, Bakhet M, Allo G, Jewett MA, Girgis AH, Latif A, Girgis H, Von Both I, Bjarnason GA, Yousef GM. miR-192, miR-194 and miR-215: a convergent microRNA network suppressing tumor progression in renal cell carcinoma. Carcinogenesis. 2013;34(10):2231-9.

http://dx.doi.org/10.1093/carcin/bgt184 PMid:23715501.

30. Ueno K, Hirata H, Shahryari V, Chen Y, Zaman MS, Singh K, Tabatabai ZL, Hinoda Y, Dahiya R. Tumour suppressor microRNA-584 directly targets oncogene Rock-1 and decreases invasion ability in human clear cell renal cell carcinoma. Br J Cancer. 2011;104(2):308-15.

http://dx.doi.org/10.1038/sj.bjc.6606028 PMid:21119662 PMCid:PMC3031891.

- 31. Yamada Y, Hidaka H, Seki N, Yoshino H, Yamasaki T, Itesako T, Nakagawa M, Enokida H. Tumor-suppressive microRNA-135a inhibits cancer cell proliferation by targeting the c-MYC oncogene in renal cell carcinoma. Cancer Sci. 2013;104(3):304-12. http://dx.doi.org/10.1111/cas.12072 PMid:23176581.
- 32. Majid S et al. MicroRNA-205 inhibits Src-mediated oncogenic pathways in renal cancer. Cancer Res. 2011;71(7):2611-21. http://dx.doi.org/10.1158/0008-5472.CAN-10-3666 PMid:21330408 PMCid:PMC3940352.
- 33. Saini S, Yamamura S, Majid S, Shahryari V, Hirata H, Tanaka Y, Dahiya R. MicroRNA-708 induces apoptosis and

suppresses tumorigenicity in renal cancer cells. Cancer Res. 2011;71(19):6208-19. http://dx.doi.org/10.1158/0008-5472.CAN-11-0073 PMid:21852381 PMCid:PMC3940359.

- 34. Hirata H, Hinoda Y, Ueno K, Nakajima K, Ishii N, Dahiya R. MicroRNA-1826 directly targets beta-catenin (CTNNB1) and MEK1 (MAP2K1) in VHL-inactivated renal cancer. Carcinogenesis. 2012;33(3):501-8. http://dx.doi.org/10.1093/carcin/bgr302 PMid:22180573 PMCid:PMC3291860.
- 35. Wu C, Jin B, Chen L, Zhuo D, Zhang Z, Gong K, Mao Z. MiR-30d induces apoptosis and is regulated by the Akt/FOXO pathway in renal cell carcinoma. Cell Signal. 2013;25(5):1212-21.

http://dx.doi.org/10.1016/j.cellsig.2013.0 1.028

PMid:23416459.

36. Ma X et al. MicroRNA-185 inhibits cell proliferation and induces cell apoptosis by targeting VEGFA directly in von Hippel-Lindau-inactivated clear cell renal cell carcinoma. Urol Oncol. 2015;33(4):169.e1-169.e11.

http://dx.doi.org/10.1016/j.urolonc.2015. 01.003

PMid:25700976.

37. Czyzyk-Krzeska MF, Meller J, Plas DR. Not all autophagy is equal. Autophagy. 2012; 8(7):1155-6.

http://dx.doi.org/10.4161/auto.20650 PMid:22647376 PMCid:PMC3429557.

38. Mikhaylova O et al. VHL-regulated MiR-204 suppresses tumor growth through inhibition of LC3B-mediated autophagy in renal clear cell carcinoma. Cancer Cell. 2012;21(4):532-546.

http://dx.doi.org/10.1016/j.ccr.2012.02.0

PMid:22516261 PMCid:PMC3331999.

39. Zheng B, Zhu H, Gu D, Pan X, Qian L, Xue B, Yang D, Zhou J, Shan Y. MiRNA-30a-mediated autophagy inhibition sensitizes renal cell carcinoma cells to sorafenib. Biochem Biophys Res Commun. 2015;459(2):234-9.

http://dx.doi.org/10.1016/j.bbrc.2015.02. 084

PMid:25712526.

40. Redova M, Svoboda M. and Slaby O. MicroRNAs and their target gene networks in renal cell carcinoma. Biochem. Biophys. Res. Commun. 2011;405:153-156.

http://dx.doi.org/10.1016/j.bbrc.2011.01.

PMid:21232526.

41. Petillo D, Kort EJ, Anema J, Furge KA, Yang XJ, Teh BT. MicroRNA profiling of human kidney cancer subtypes. Int J Oncol. 2009;35(1):109-14.

http://dx.doi.org/10.3892/ijo\_00000318 PMid:19513557.

42. Youssef YM, White NM, Grigull J, Krizova A, Samy C, Mejia-Guerrero S, Evans A, Yousef GM. Accurate molecular classification of kidney cancer subtypes using microRNA signature. Eur Urol. 2011;59(5):721-30.

http://dx.doi.org/10.1016/j.eururo.2011.0 1.004

PMid:21272993.

43. Spector Y, Fridman E, Rosenwald S, Zilber S, Huang Y, Barshack I, Zion O, Mitchell H, Sanden M, Meiri E. Development and validation of a microRNA-based diagnostic assay for classification of renal cell carcinomas. Mol Oncol. 2013;7(3):732-8.

http://dx.doi.org/10.1016/j.molonc.2013. 03.002

PMid:23587442.

44. Iwamoto H, Kanda Y, Sejima T, Osaki M, Okada F, Takenaka A. Serum miR-210 as a potential biomarker of early clear cell renal cell carcinoma. Int J Oncol. 2014;44(1):53-8

PMid:24212760.

45. Zhao A, Li G, Péoc'h M, Genin C, Gigante M. Serum miR-210 as a novel biomarker for molecular diagnosis of clear cell renal cell carcinoma. Exp Mol Pathol. 2013;94(1):115-20.

http://dx.doi.org/10.1016/j.yexmp.2012.1 0.005

PMid:23064048.

46. Redova M, Poprach A, Nekvindova J, Iliev R, Radova L, Lakomy R, Svoboda M, Vyzula R, Slaby O. Circulating miR-378 and miR-451 in serum are potential biomarkers

for renal cell carcinoma. J Transl Med. 2012;10:55.

http://dx.doi.org/10.1186/1479-5876-10-55

PMid:22440013 PMCid:PMC3340316.

47. Wulfken LM et al. MicroRNAs in renal cell carcinoma: diagnostic implications of serum miR-1233 levels. PLoS One. 2011;6(9):e25787.

http://dx.doi.org/10.1371/journal.pone.00 25787

PMid:21984948 PMCid:PMC3184173.

48. Wang C et al. A panel of five serum miRNAs as a potential diagnostic tool for early-stage renal cell carcinoma. Sci Rep. 2015;5:7610.

http://dx.doi.org/10.1038/srep07610 PMid:25556603.

49. McCormick RI, Blick C, Ragoussis J, Schoedel J, Mole DR, Young AC, Selby PJ, Banks RE, Harris AL. miR-210 is a target of hypoxia-inducible factors 1 and 2 in renal cancer, regulates ISCU and correlates with good prognosis. Br J Cancer. 2013;108(5):1133-42.

http://dx.doi.org/10.1038/bjc.2013.56 PMid:23449350 PMCid:PMC3619073.

50. Samaan S et al. miR-210 is a prognostic marker in clear cell renal cell carcinoma. J Mol Diagn. 2015;17(2):136-44.

http://dx.doi.org/10.1016/j.jmoldx.2014.1 0.005

PMid:25555365.

51. Khella HW et al. Low expression of miR-126 is a prognostic marker for metastatic clear cell renal cell carcinoma. Am J Pathol. 2015;185(3):693-703.

http://dx.doi.org/10.1016/j.ajpath.2014.1 1.017

PMid:25572155.

52. Vergho D, Kneitz S, Rosenwald A, Scherer C, Spahn M, Burger M, Riedmiller H, Kneitz B. Combination of expression levels of miR-21 and miR-126 is associated with cancer-specific survival in clear-cell renal cell carcinoma. BMC Cancer. 2014;14:25.

http://dx.doi.org/10.1186/1471-2407-14-25

PMid:24428907 PMCid:PMC3897948.

53. Fu Q, Liu Z, Pan D, Zhang W, Xu L, Zhu Y, Liu H, Xu J. Tumor miR-125b predicts recurrence and survival of patients with clear-cell renal cell carcinoma after surgical resection. Cancer Sci. 2014;105(11):1427-34.

http://dx.doi.org/10.1111/cas.12507 PMid:25155155.

54. Fritz HK, Lindgren D, Ljungberg B, Axelson H, Dahlbäck B. The miR(21/10b) ratio as a prognostic marker in clear cell renal cell carcinoma. Eur J Cancer. 2014;50(10):1758-65.

http://dx.doi.org/10.1016/j.ejca.2014.03. 281

PMid:24793999.

55. Heinzelmann J, Unrein A, Wickmann U, Baumgart S, Stapf M, Szendroi A, Grimm MO, Gajda MR, Wunderlich H, Junker K. MicroRNAs with prognostic potential for metastasis in clear cell renal cell carcinoma: a comparison of primary tumors and distant metastases. Ann Surg Oncol. 2014;21(3):1046-54.

http://dx.doi.org/10.1245/s10434-013-3361-3

PMid:24242678.

56. Gowrishankar B, Ibragimova I, Zhou Y, Slifker MJ, Devarajan K, Al-Saleem T, Uzzo RG, Cairns P. MicroRNA expression signatures of stage, grade, and progression in clear cell RCC. Cancer Biol Ther. 2014;15(3):329-41.

http://dx.doi.org/10.4161/cbt.27314 PMid:24351440 PMCid:PMC3974834.

57. Ge YZ et al. A tumor-specific microRNA signature predicts survival in clear cell renal cell carcinoma. J Cancer Res Clin Oncol. 2015;141(7):1291-9.

http://dx.doi.org/10.1007/s00432-015-1927-0

- 58. Chang I et al. Loss of miR-200c upregulates CYP1B1 and confers docetaxel resistance in renal cell carcinoma. Oncotarget. 2015;6(10):7774-87. PMid:25860934 PMCid:PMC4480715.
- 59. Gao C, Peng FH, Peng LK. MiR-200c sensitizes clear-cell renal cell carcinoma cells to sorafenib and imatinib by targeting heme oxygenase-1. Neoplasma. 2014;61(6):680-9.

http://dx.doi.org/10.4149/neo\_2014\_083 PMid:25150313.

60. Prior C et al. Identification of tissue microRNAs predictive of sunitinib activity in patients with metastatic renal cell carcinoma. PLoS One. 2014;9(1):e86263. http://dx.doi.org/10.1371/journal.pone.00 86263

PMid:24475095 PMCid:PMC3901669.

61. Berkers J et al. A possible role for microRNA-141 down-regulation in sunitinib resistant metastatic clear cell renal cell carcinoma through induction of epithelial-to-mesenchymal transition and hypoxia resistance. J Urol. 2013;189(5):1930-8. http://dx.doi.org/10.1016/j.juro.2012.11. 133

PMid:23206420.

62. Gámez-Pozo A et al. MicroRNA expression profiling of peripheral blood samples predicts resistance to first-line sunitinib in advanced renal cell carcinoma patients. Neoplasia. 2012 Dec;14(12):1144-52

http://dx.doi.org/10.1593/neo.12734 PMid:23308047 PMCid:PMC3541518.

- 63. Pecot CV et al. Tumour angiogenesis regulation by the miR-200 family. Nat Commun. 2013;4:2427.
- http://dx.doi.org/10.1038/ncomms3427
- 64. Cui L et al. MicroRNA-99a induces G1-phase cell cycle arrest and suppresses tumorigenicity in renal cell carcinoma. BMC Cancer. 2012;12:546.

http://dx.doi.org/10.1186/1471-2407-12-546

- 65. Ben-Shushan D, Markovsky E, Gibori H, Tiram G, Scomparin A, Satchi-Fainaro R. Overcoming obstacles in microRNA delivery towards improved cancer therapy. Drug Deliv Transl Res. 2014;4(1):38-49. http://dx.doi.org/10.1007/s13346-013-0160-0
- 66. Maroto P, Rini B. Molecular biomarkers in advanced renal cell carcinoma. Clin Cancer Res. 2014;20(8):2060-71. http://dx.doi.org/10.1158/1078-0432.CCR-13-1351 PMid:24526734.